

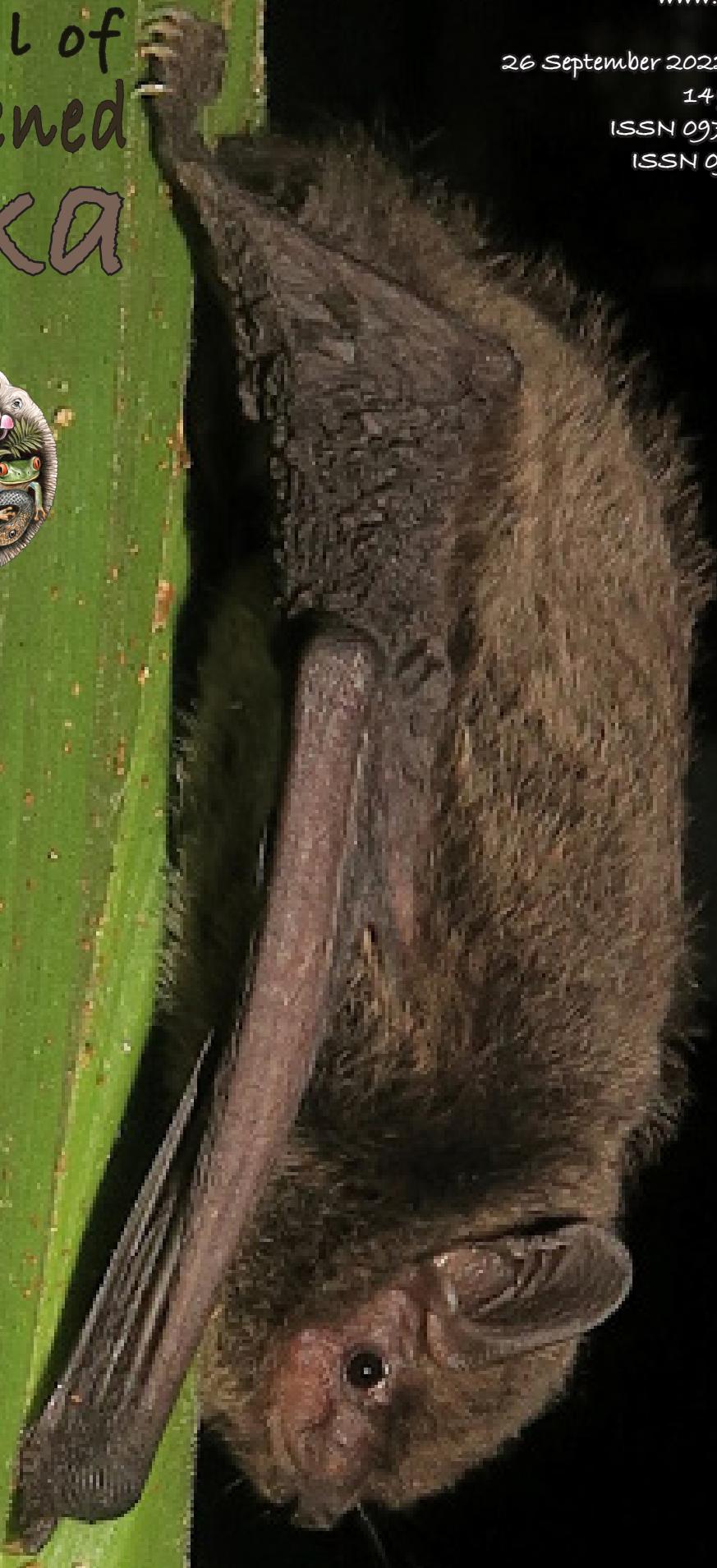
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Cover: *Pipistrellus tenuis* recorded during the small mammalian fauna study, Manipur, India. © Uttam Saikia.



Invasive alien plant species of Hassan District, Karnataka, India

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Abstract: This study was undertaken to document alien and invasive flowering plant species in the Western Ghats (Hassan district, Karnataka, India), with background information on family, habit, habitat, longevity, nativity, and uses. A total of 312 alien species belonging to 236 genera in 79 families are listed. The majority belong to family Asteraceae (36 species), followed by Fabaceae (21 species), and Amaranthaceae (17 species). Herbs constitute the majority (59%) of alien species followed by shrubs (17%). Around 36% the alien taxa are native to tropical America. Of 314 alien species, 122 were intentional introductions, with a majority (39%) introduced for ornamental purposes; 24% of species have naturalized, while 33% display as invasive. There is an urgent need to gather regional data on the diversity of invasive alien plant species in order to study the impact on native vegetation and biodiversity.

Keywords: Exotic, naturalized species, ornamental, plant diversity, threats, Western Ghats.

Editor: Anonymity requested.

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Author contribution: GMP carried out the floristic study, collected the data and wrote the manuscript. SN identified the species, interpreted the data and designed the manuscript. Both authors have read and approved the final manuscript.

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INTRODUCTION

The increase in human activity and increased international trade, travel, and transport beyond biogeographic barriers has led to the introduction and establishment of invasive alien species in new regions (Dawson et al. 2017). Biological invasions have received much consideration due to the potential threats they impose on native species, natural systems, ecosystem processes & functioning, environmental quality, and human health (Pyšek & Richardson 2010; Simberloff et al. 2013; Jones & McDermott 2018; Pearson et al. 2018; Petruzzella et al. 2018; Bartz & Kowarik 2019; Rai & Singh 2020). Successful plant invasions are attributed to the interaction between the exotic plants and resident plant communities (Gallien & Carboni 2017). Many factors influence invasion success, including phenotypic plasticity, dispersal benefits from destructive foraging activities, wide geographic range, vegetative reproduction, fire tolerance, and superior competitive ability compared to native flora (Sharma et al. 2005). Invasive alien plants may outperform native species due to the absence of natural enemies in the introduced range (Aguilera 2011). Moreover, invasive plants display characteristics such as high competitive ability and efficient resource utilization (Baker 1965; Levine 2000; Petruzzella et al. 2018).

The introduction of non-native species into new habitats is largely due to short-term economic benefits (MeNeely 2001), therefore, most of the issues related to invasive plants can be linked to the intended or unintended consequences of economic activities (Perrings et al. 2002). Globalization and rapid modification of natural habitats have triggered a massive spread of plant species to areas outside their native ranges (van Kleunen et al. 2015). On the continental and global scale, species invasions have diminished the regional distinctiveness of flora and fauna (Vitousek et al. 1997). At least 10% of the world's vascular plants (300,000) have the potential to invade other ecosystems and affect native biota in direct or indirect ways (Singh et al. 2006). About 18% of the Indian flora are aliens, of which 55% are native to the Americas, 30% to Asia, and to 15% Europe & central Asia (Nayar 1977; Singh et al. 2010). Many invasive alien plants confer economic benefits; for example, *Lantana camara* is used by several local villages in India who use it for furniture and pulp making (Kannan et al. 2014); however, it remains a serious invader causing problems for indigenous flora and significant losses of ecosystem services compared to benefits.

Many invasive species have severe negative impacts.

For example, *Ageratum conyzoides*, *Chromolaena odorata*, and *Parthenium hysterophorus* are considered invasive transformer species that lack natural enemies and have fast-spreading ability, allelopathic effects on other plants, and strong competitiveness with crops, while posing health hazard to humans and animals (Raghubanshi et al. 2005; Suthari et al. 2016). Some cultivated alien species provide food, medicine, fuel, & fodder to local communities (Kull et al. 2007) and some are used in the preparation of Ayurvedic formulations (Shiddamallayya et al. 2010). It is estimated that as many as 50% of invasive species, in general, can be classified as ecologically harmful, based on their actual impacts (Richardson et al. 2000).

There is a need for an authoritative database on alien and invasive alien plant species to monitor the spread and impact in various regions and for plan appropriate management strategies. State and regional floras in the country rarely indicate the native or alien status of the species listed therein. In some cases, naturalized alien species are treated as native in floristic documents (Khuroo et al. 2012). Many species recorded as aliens in different regions of the country, but whose native range falls within the country's political boundary, have been excluded in the present study. Example is the Himalayan Chir Pine *Pinus roxburghii* recorded as 'exotic' in southern India (Matthew 1969). Similarly, *Nyctanthes arbor-tristis* is a Himalayan native introduced for various reasons to the rest of the country, and many other species that have a within-the-country origin should also be regarded as alien. The present study reports on alien and invasive flowering plant species in the Hassan district of Karnataka.

MATERIALS AND METHODS

Study area

Hassan district is located in the southern part of Karnataka state in India, situated between 12° 13', 13° 33' N & 75° 33', 76° 38' E. Hassan district begins at the base of the steep Western Ghats and continues into the gently rolling Deccan plateau. The district shows wide variations in climate and vegetation. The evergreen and semi-evergreen forests in the district are concentrated in the Western Ghats region of Yeslur and Sakaleshpura, and are commonly known as wet evergreen tropical rain forest. Dry deciduous forests dominate the plains, also known as Maidan area (Figure 1).

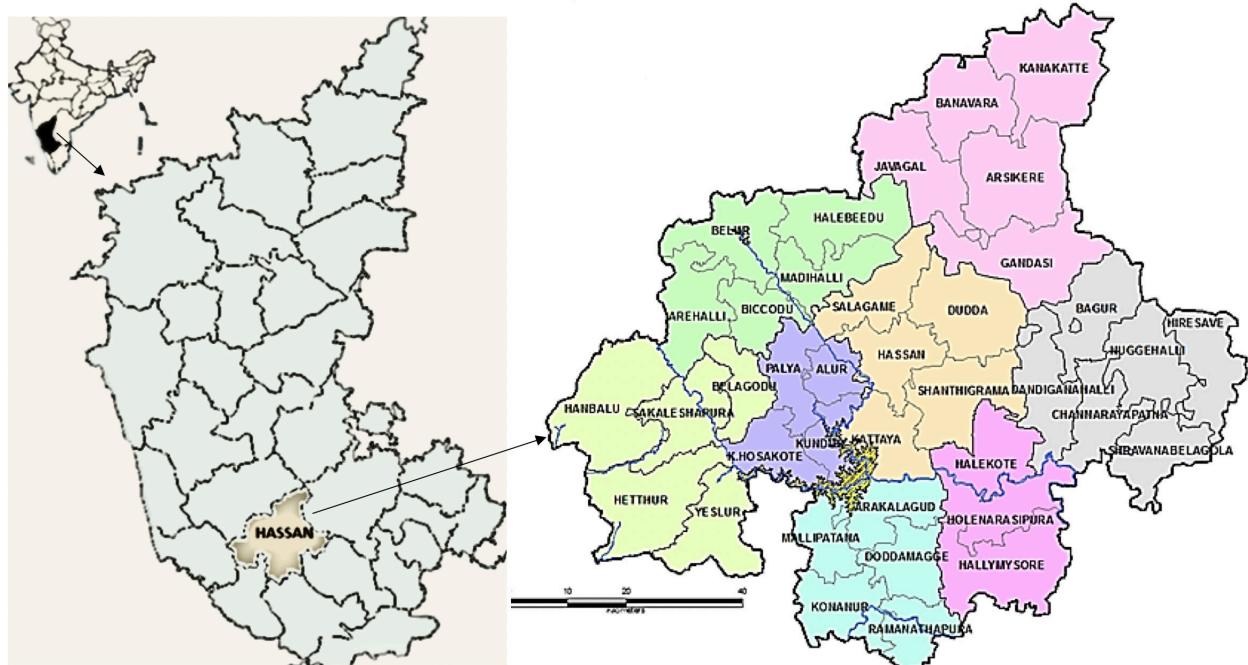


Figure 1. The study area of Hassan District of Karnataka.

Data collection

Extensive floristic surveys were conducted in a planned manner repeatedly in different seasons to get the maximum representation of alien and invasive alien species in Hassan District. Plant samples were collected from natural habitats, agricultural lands, aquatic, semi-aquatic habitats, marshes, open grasslands, wastelands, roadsides, village ponds, wetlands, railway tracks, riverbanks, reserve forests, slopes, and hilltops. The collected specimens were identified with the help of floras (Saldhana & Nicolson 1976; Saldhana 1984, 1996). Plants were categorized by habit (herb, shrub, climber, and tree) and by habitat (wasteland, cultivated field, aquatic, river & pond banks, forest, and roadside). The plant names were rechecked and authenticated using the plant list ([www.theplantlist.org\](http://www.theplantlist.org/)) and GRIN taxonomy site (<http://www.ars-grin.gov/npgs/aboutgrin.html>), the synonyms were removed to avoid taxonomic inflation. We followed biogeographic approach in assigning the native ranges to all the species (Khuroo et al. 2012). Only those species whose native ranges fall outside the borders of the Indian subcontinent, namely 'alien' species (CBD 2000) were considered in this study. To minimize the error of judgement by earlier studies regarding the alien status, and to cross-check native range records, native ranges for all species were verified with data from the Germplasm Resources Information Network (www.grin.org, <http://www.hear.org/pier/>, <http://www.iucngisd.org/gis/>) and some other published literature (Murthy et al. 2007; Negi & Hajra 2007; Reddy 2008; Reddy et al. 2008; Singh et al. 2010; Wu et al. 2010; Paul 2010; Khuroo et al. 2012; Pyšek et al. 2012).

To further document their status, alien plant species were categorized into casual (Ca), naturalized (Nt), invasive (In), casual or naturalized (Ca/Nt) and naturalized or invasive (Nt/In) as per the earlier studies (Richardson et al. 2000; Pysek et al. 2004; Khuroo et al. 2012). Alien species that may flourish and even reproduce occasionally in an area, but do not form self-replacing populations, and which rely on repeated introductions for their persistence are known as 'casual' (Ca). Alien species that reproduce and sustain populations over more than one life cycle and do not necessarily invade natural, semi-natural or human-made ecosystems are known as 'naturalized' (Nt). Naturalized alien species that produce reproductive offspring, often in large numbers, at considerable distances from parent plants and thus can spread over a considerable area are referred to as 'invasive' (In). Alien species grown or planted and have not yet escaped are referred to as 'cultivated' (Cl). Those casual alien species for which the current evidence is insufficient to be recognized as naturalized but have the potential to become naturalized in the near future are referred to as Ca/Nt. Those naturalized alien species for which the current evidence is insufficient to be recognized as invasive, but have the potential to become

invasive in the near future are referred to as Nt/In. The purpose of intentional introduction (food, fodder, ornamental, plantation, horticulture, and medicinal) of the alien species were recorded from relevant literature (Sharma & Pandey 1984; Khuroo et al. 2007; Jaryan et al. 2013). Species that have come unintentionally were categorized as 'unintentional introductions' (Ui). Literature including unpublished (Singh et al. 2010; Kambhar & Kotresha 2011; Prakash & Balasubramanian 2018) and local communities were consulted for uses. The alien species were analyzed for taxa statistics, habit, habitat, nativity, purpose of introduction, invasive status and use-values. For analysis of habit, the number of species in a particular habit has been divided by the total number of alien species and multiplied by 100. The same follows for habitat, nativity, and invasion status analyses. For analyzing the purpose of introduction, number of species introduced for a particular purpose was divided by the total number of species for which the purpose of introduction is known (122) and then multiplied by hundred. We used Microsoft Excel (version 2013) for the data processing.

RESULTS AND DISCUSSION

A total of 312 species in 236 genera and under 79 families were documented as invasive alien plant species. They are shown along with the family name, habit, habitat, nativity, mode of introduction, invasive status, and uses in Table 1 and Images 1–5. The habit-wise distribution of alien species is represented in Figure 2.

Of the species 48% ($n = 152$) belong to just 10 families. Asteraceae was the dominant family with 36 species (23%) followed by Fabaceae 21 species (13%), Amaranthaceae 17 species (11%), and Poaceae 16 species (10%). Due to their dominance, most of these families have a high number of herbs. The dominance of Fabaceae, which has the ability to fix nitrogen, would aid their colonisation of empty niches. The proportion of alien species to the total species in the respective family in Hassan district is highest for Amaranthaceae (85%) followed by Solanaceae (83.3%) and Asteraceae (47.3%) (Table 2). For India, this is in agreement with Khuroo et al. (2012) and Jaryan et al. (2013). Similar patterns of family dominance in alien floras have been reported in studies from Europe (Lambdon et al. 2008) and China (Weber et al. 2008; Wu et al. 2010). Pysek (1998) found that these families also have the majority of alien species on a worldwide scale. In addition, studies on agricultural

weeds found that the Asteraceae and Poaceae families account for the majority of weeds in terms of numbers (Heywood 1989). This could be due to the fact that these families have some of the highest species richness (Rao 1994) and hence have a higher chance of harboring more alien species. Such a family dominance pattern, as Khuroo et al. (2012) pointed out, is more of a depiction of sampling effect. These families are known to have a large number of species, hence an increase in the number of alien species belonging to these families is expected (Khuroo et al. 2012). In Himachal Pradesh also, proportion of alien species relative to the total species in Amaranthaceae (53.3%) followed by Solanaceae (52.9%) and Convolvulaceae (44%) is highest (Jaryan et al. 2013). This is in agreement to the results of Khuroo et al. (2012) for India and Wu et al. (2010) for China. In Kashmir, proportion of alien species relative to the total species in Amaranthaceae (83%) is highest (Khuroo et al. 2007). Interestingly, the Asteraceae and Convolvulaceae families have the biggest numerical contributions (47.3%) in this ranking for Hassan district (Table 2). In the top 10 list of families, Poaceae (11.4%) has the lowest alien species (Table 2). Remarkably, some families comprise only invasive species in Hassan district (e.g., Balsaminaceae, *Impatiens balsamina*; Ceratophyllaceae, *Ceratophyllum demersum*; Martyniaceae, *Martynia annua*). The following genera had highest number of alien species in Hassan district, *Solanum* (8 species), *Ipomoea* (7 species), and *Euphorbia* (6 species), *Amaranthus*, *Alternanthera*, & *Hibiscus* (4 species each). These genera also contribute a good number to the alien flora of India, Europe, and China (Lambdon et al. 2008; Weber et al. 2008; Wu et al. 2010; Khuroo et al. 2012).

Out of the 36 plant species that are globally recognised as the 'World's worst invasive alien species' (Lowe et al. 2000), 17 are present in India (Khuroo et al. 2012), of which we report the presence of eight in Hassan district: *Eichhornia crassipes*, *Spathodea campanulata*, *Imperata*

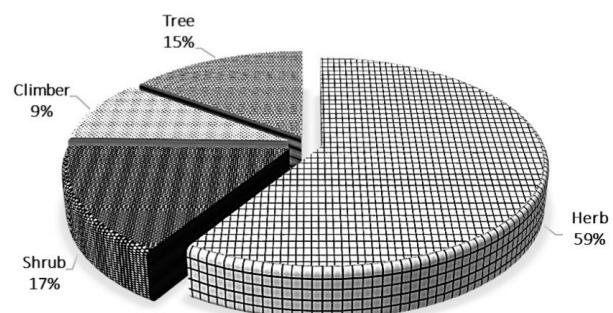


Figure 2. Habit-wise distribution of alien species of Hassan District.

Table 1. Alien plant species of Hassan District, their source region and uses.

| | Accepted name of species | Family | Habit | Longevity | Habitat | Nativity | Purpose of introduction | Invasive status | Uses |
|----|---|------------------|-------|-----------|---------|----------|-------------------------|-----------------|------|
| 1 | <i>Abelmoschus esculentus</i> (L.) Moench | Malvaceae | H | A | CF | TAF | Fd | Cl | V |
| 2 | <i>Acacia auriculiformis</i> L. | Mimosaceae | T | P | AR | AU | Ui | Ca/Nt | W |
| 3 | <i>Acacia farnesiana</i> (L.) Willd. | Mimosaceae | T | P | AR | SAM | Ui | In | M |
| 4 | <i>Acanthospermum hispidum</i> DC. | Asteraceae | H | A | W | BR | Ui | In | M |
| 5 | <i>Achyranthes aspera</i> L. | Amaranthaceae | H | A | W | AS | Ui | In | M |
| 6 | <i>Achyranthes bidentata</i> Blume | Amaranthaceae | H | P | AR | AS | Ui | Nt | M |
| 7 | <i>Acmella uliginosa</i> (Sw.) Cass. | Asteraceae | H | A | W | TAM | Ui | Nt | Nk |
| 8 | <i>Adenostemma lavenia</i> (L.) Ktze. | Asteraceae | H | A | RB | SAM | Ui | In | Nk |
| 9 | <i>Aeschynomene indica</i> L. | Fabaceae | H | A | AQ | AU | Ui | In | Nk |
| 10 | <i>Agave americana</i> L. | Asparagaceae | S | P | AR | TAM | Ui | Nt | R |
| 11 | <i>Agave sisalana</i> Perrine. | Asparagaceae | S | P | W | MX | Ui | Cl | R |
| 12 | <i>Ageratum conyzoides</i> (L.) L. | Asteraceae | H | A | W | TAM | Or | In | M |
| 13 | <i>Ageratum houstonianum</i> Mill. | Asteraceae | H | A | W | TAM | Ui | In | Nk |
| 14 | <i>Albizia lebbeck</i> Benth. | Mimosaceae | T | P | F | AS | Pl | Nt | W |
| 15 | <i>Albizia saman</i> (Jacq.) Merr. | Mimosaceae | T | P | AR | TAM | Ui | Ca/Nt | W |
| 16 | <i>Allamanda cathartica</i> L. | Apocynaceae | C | P | CF | TAM | Or | Cl | Or |
| 17 | <i>Allium cepa</i> L. | Amaryllidaceae | H | A | CF | AS | Fd | Cl | V |
| 18 | <i>Allium sativum</i> L. | Amaryllidaceae | H | A | CF | AS | Fd | Cl | V |
| 19 | <i>Aloe vera</i> (L.) Burm.f. | Liliaceae | H | P | W | MR | M | Ca/Nt | M |
| 20 | <i>Alternanthera paronychioides</i> A.St.-Hil. | Amaranthaceae | H | P | RB | TAM | Ui | Nt/In | M |
| 21 | <i>Alternanthera philoxeroides</i> (Mart.) Griseb. | Amaranthaceae | H | P | W | TAM | Ui | Nt/In | Nk |
| 22 | <i>Alternanthera pungens</i> Kunth | Amaranthaceae | H | P | W | TAM | Ui | Nt/In | M |
| 23 | <i>Alternanthera sessilis</i> (L.) R.Br. ex DC. | Amaranthaceae | H | P | RB | TAM | Ui | Nt | V |
| 24 | <i>Amaranthus caudatus</i> L. | Amaranthaceae | H | A | CF | SAM | Fd | In | V |
| 25 | <i>Amaranthus spinosus</i> L. | Amaranthaceae | H | A | CF | TAM | Ui | In | V |
| 26 | <i>Amaranthus tricolor</i> L. | Amaranthaceae | H | A | CF | AS | Fd | Ca | V |
| 27 | <i>Amaranthus viridis</i> L. | Amaranthaceae | H | A | CF | TAM | Ui | In | V |
| 28 | <i>Ammannia baccifera</i> L. | Lythraceae | H | A | RB | AU | Ui | Nt | Nk |
| 29 | <i>Anacardium occidentale</i> L. | Anacardiaceae | T | P | W | BR | Ht | Nt | M |
| 30 | <i>Anagallis arvensis</i> L. | Primulaceae | H | A | RB | EU | Ui | In | Nk |
| 31 | <i>Ananas comosus</i> (L.) Merr. | Bromeliaceae | H | P | CF | SAM | Ht | Cl | Ef |
| 32 | <i>Anethum graveolens</i> L. | Apiaceae | H | A | CF | AS | Or | Nt | V |
| 33 | <i>Annona muricata</i> L. | Annonaceae | T | P | CF | TAM | Ht | Cl | Ef |
| 34 | <i>Annona reticulata</i> L. | Annonaceae | T | P | F | TAM | Ht | Cl | Ef |
| 35 | <i>Annona squamosa</i> L. | Annonaceae | T | P | F | WI | Ht | Cl | Ef |
| 36 | <i>Antigonon leptopus</i> Hook. & Arn. | Polygonaceae | C | P | AR | TAM | Or | Ca/Nt | Or |
| 37 | <i>Arachis hypogaea</i> L. | Fabaceae | H | A | CF | BR | Fd | Cl | Ol |
| 38 | <i>Areca catechu</i> L. | Arecaceae | T | P | CF | AS | Pl | Nt | En |
| 39 | <i>Argemone mexicana</i> L. | Papaveraceae | H | A | W | NAM | M | In | M |
| 40 | <i>Aristolochia littoralis</i> Parodi | Aristolochiaceae | H | P | W | BR | Ui | Cl | Or |
| 41 | <i>Arthraxon lancifolius</i> (Trin.) Hochst. | Poaceae | H | A | W | TAF | Ui | Nt | V |
| 42 | <i>Artocarpus altilis</i> (Parkinson ex F.A.Zorn) Fosberg | Moraceae | T | P | W | SEA | Fd | Cl | Ef |
| 43 | <i>Asclepias curassavica</i> L. | Apocynaceae | H | P | AR | TAM | Ui | Ca/Nt | Nk |
| 44 | <i>Averrhoa bilimbi</i> L. | Oxalidaceae | T | P | AR | TAM | Ui | Cl | Ef |

| | Accepted name of species | Family | Habit | Longevity | Habitat | Nativity | Purpose of introduction | Invasive status | Uses |
|----|--|------------------|-------|-----------|---------|----------|-------------------------|-----------------|------|
| 45 | <i>Averrhoa carambola</i> L. | Oxalidaceae | T | P | AR | TAM | Ui | Cl | Ef |
| 46 | <i>Bacopa monnieri</i> Pennell | Scrophulariaceae | H | A | RB | TAM | Ui | In | V |
| 47 | <i>Balanites aegyptiaca</i> (L.) Delile | Zygophyllaceae | S | P | F | TAF | Ui | Nt | M |
| 48 | <i>Bambusa vulgaris</i> Schrad. | Poaceae | S | P | AR | SEA | Or | Ca/Nt | Or |
| 49 | <i>Basella alba</i> L. | Basellaceae | C | A | CF | TAF | M | Nt | M |
| 50 | <i>Benincasa hispida</i> (Thunb.) Cogn. | Cucurbitaceae | C | A | CF | SEA | Fd | Cl | V |
| 51 | <i>Beta vulgaris</i> L. | Amaranthaceae | H | A | CF | EU | Ht | Cl | V |
| 52 | <i>Bidens biternata</i> (Lour.) Merr. & Sheriff | Asteraceae | H | A | CF | TAM | Ui | Ca/Nt | M |
| 53 | <i>Bidens pilosa</i> L. | Asteraceae | H | A | W | SAM | Ui | In | Nk |
| 54 | <i>Biophytum sensitivum</i> DC. | Oxalidaceae | H | A | W | SEA | Ui | In | M |
| 55 | <i>Bixa orellana</i> L. | Bixaceae | T | P | CF | BR | Ui | Cl | M |
| 56 | <i>Blainvillea acmella</i> (L.) Philipson | Asteraceae | H | A | W | TAM | Ui | In | M |
| 57 | <i>Blumea lacera</i> (Burm.f.) DC. | Asteraceae | H | A | W | TAM | Ui | In | M |
| 58 | <i>Blumea obliqua</i> (L.) Druce | Asteraceae | H | A | W | TAM | Ui | In | Nk |
| 59 | <i>Bougainvillea spectabilis</i> Willd. | Nyctaginaceae | S | P | AR | TAM | Or | Cl | Or |
| 60 | <i>Brassica nigra</i> (L.) K.Koch | Brassicaceae | H | A | CF | EU | Fd | Cl | Ol |
| 61 | <i>Brassica oleracea</i> L. | Brassicaceae | H | A | CF | EU | Fd | Cl | V |
| 62 | <i>Breynia vitis-idea</i> (Burn.f.) Fisch | Euphorbiaceae | S | A | CF | WI | Ui | Nt | Nk |
| 63 | <i>Brugmansia suaveolens</i> Bercht. & K.Presl. | Solanaceae | S | P | AR | BR | UI | Nt | Nk |
| 64 | <i>Bryophyllum pinnatum</i> (Lam.) Oken | Crassulaceae | H | A | W | TAF | Ui | Nt/In | M |
| 65 | <i>Caesalpinia pulcherrima</i> (L.) Sw. | Caesalpiniaceae | S | P | CF | TAM | Or | Cl | Or |
| 66 | <i>Cajanus cajan</i> (L.) Millsp. | Fabaceae | H | A | CF | TAF | Fd | Cl | ES |
| 67 | <i>Caladium bicolor</i> (Aiton) Vent. | Araceae | H | A | RB | TAM | Or | Cl | Or |
| 68 | <i>Callistemon viminalis</i> (Sol. ex Gaertn.) G.Don ex Loudon | Myrtaceae | T | P | CF | AU | Or | Ca/Nt | Or |
| 69 | <i>Calotropis gigantea</i> (L.) Dryand. | Apocynaceae | S | P | W | TAF | Ui | In | M |
| 70 | <i>Camellia sinensis</i> (L.) Kuntze | Theaceae | S | P | CF | AS | Fd | Ca/Nt | Br |
| 71 | <i>Canna indica</i> L. | Cannaceae | H | P | CF | TAM | M | Nt | M |
| 72 | <i>Capsicum annuum</i> L. | Solanaceae | H | P | CF | MX | Ht | Cl | V |
| 73 | <i>Cardiospermum halicacabum</i> L. | Sapindaceae | C | P | W | SAM | Ui | In | M |
| 74 | <i>Carica papaya</i> L. | Caricaceae | T | P | CF | SAM | Ht | Cl | Ef |
| 75 | <i>Carmona retusa</i> (Vahl) Masamune | Boraginaceae | H | A | F | SEA | Ui | Ca/Nt | M |
| 76 | <i>Cascabela thevetia</i> (L.) Lippold | Apocynaceae | T | P | CF | TAM | Or | Cl | Or |
| 77 | <i>Cassia fistula</i> L. | Caesalpiniaceae | T | P | F | AS | M | Nt | M |
| 78 | <i>Cassytha filiformis</i> L. | Lauraceae | C | P | P | AU | Ui | Nt | M |
| 79 | <i>Casuarina equisetifolia</i> L. | Casuarinaceae | T | P | CF | TAM | Ui | Nt | W |
| 80 | <i>Catharanthus pusillus</i> (Murray) G.Don | Apocynaceae | H | A | CF | TAM | Or | In | M |
| 81 | <i>Catharanthus roseus</i> (L.) G.Don | Apocynaceae | H | A | W | TAM | Or | In | M |
| 82 | <i>Celosia argentea</i> L. | Amaranthaceae | H | A | CF | TAM | Fd | Ca | V |
| 83 | <i>Ceratophyllum demersum</i> L. | Ceratophyllaceae | H | P | AQ | NAM | Ui | In | M |
| 84 | <i>Cereus repandus</i> (L.) Mill. | Cactaceae | S | P | AR | TAM | Ui | Nt | M |
| 85 | <i>Cestrum nocturnum</i> L. | Solanaceae | S | P | AR | WI | Or | Ca/Nt | Or |
| 86 | <i>Chenopodium album</i> L. | Amaranthaceae | H | A | CF | EU | Fd | In | V |
| 87 | <i>Chenopodium ambrosioides</i> L. | Amaranthaceae | H | A | W | TAM | Ui | In | Fo |
| 88 | <i>Chloris barbata</i> Sw. | Poaceae | H | A | W | TAM | Ui | Nt | Fo |
| 89 | <i>Chromolaena odorata</i> (L.) RM.King & H.Rob. | Asteraceae | H | P | W | TAM | Ui | In | M |

| | Accepted name of species | Family | Habit | Longevity | Habitat | Nativity | Purpose of introduction | Invasive status | Uses |
|-----|--|-----------------|-------|-----------|---------|----------|-------------------------|-----------------|------|
| 90 | <i>Cicer arietinum</i> L. | Fabaceae | H | A | CF | AS | Fd | Cl | Es |
| 91 | <i>Citrullus lanatus</i> (Thunb.) Matsum. & Nakai | Cucurbitaceae | C | A | CF | SAM | Fd | Cl | Ef |
| 92 | <i>Cleome monophylla</i> L. | Cleomaceae | H | A | AR | TAF | Ui | Nt | M |
| 93 | <i>Cleome viscosa</i> L. | Cleomaceae | H | A | W | TAM | Ui | Nt | M |
| 94 | <i>Clidemia hirta</i> (L.) D. Don | Melastomataceae | H | P | W | TAM | Ui | Nt | M |
| 95 | <i>Clitoria ternatea</i> L. | Fabaceae | C | A | W | TAM | M | Nt | M |
| 96 | <i>Coffea arabica</i> L. | Rubiaceae | S | P | CF | TAF | Fd | Cl | Br |
| 97 | <i>Coldenia procumbens</i> L. | Boraginaceae | H | A | W | NAM | Ui | Nt/In | M |
| 98 | <i>Colocasia esculenta</i> (L.) Schott | Araceae | H | A | RB | AS | UI | Nt | V |
| 99 | <i>Corchorus aestuans</i> L. | Malvaceae | H | A | W | TAM | Ui | Nt | M |
| 100 | <i>Corchorus trilocularis</i> L. | Malvaceae | H | A | W | TAF | Ui | In | M |
| 101 | <i>Coriandrum sativum</i> L. | Apiaceae | H | A | CF | AS | Fd | Cl | V |
| 102 | <i>Cosmos bipinnatus</i> Cav. | Asteraceae | H | A | CF | TAM | Or | Nt/In | Nk |
| 103 | <i>Couroupita guianensis</i> Aubl. | Lecythidaceae | T | P | AR | SAM | Ui | Cl | M |
| 104 | <i>Crassocephalum crepidioides</i> (Benth.) S.Moore | Asteraceae | H | A | F | TAM | Ui | In | Nk |
| 105 | <i>Crotalaria pallida</i> Aiton | Fabaceae | H | A | CF | TAM | Ui | Nt | Nk |
| 106 | <i>Crotalaria retusa</i> L. | Fabaceae | H | A | CF | TAM | Ui | Nt | Nk |
| 107 | <i>Croton bonplandianus</i> Baill. | Euphorbiaceae | H | P | W | SAM | Ui | In | Nk |
| 108 | <i>Cucumis melo</i> L. | Cucurbitaceae | C | A | CF | AS | Fd | Cl | Ef |
| 109 | <i>Cucurbita maxima</i> Duchesne | Cucurbitaceae | C | A | CF | SAM | Fd | Cl | V |
| 110 | <i>Cucurbita pepo</i> L. | Cucurbitaceae | C | A | CF | SAM | Fd | Cl | V |
| 111 | <i>Cuscuta reflexa</i> Roxb. | Convolvulaceae | C | P | P | MR | Ui | In | M |
| 112 | <i>Cyanthillium cinereum</i> (L.) H.Rob. | Asteraceae | H | A | W | AS | Ui | Nt/In | M |
| 113 | <i>Cymbopogon citratus</i> (DC.) Stapf | Poaceae | H | A | CF | SEA | Ui | Ca | Ol |
| 114 | <i>Cyperus difformis</i> L. | Cyperaceae | H | A | CF | TAM | Ui | In | Nk |
| 115 | <i>Cyperus iria</i> L. | Cyperaceae | H | A | CF | TAM | Ui | Nt | Nk |
| 116 | <i>Cyperus rotundus</i> L. | Cyperaceae | H | A | CF | TAF | Ui | In | M |
| 117 | <i>Datura metel</i> L. | Solanaceae | S | P | W | TAM | Ui | Ca/Nt | M |
| 118 | <i>Datura stramonium</i> L. | Solanaceae | S | P | AR | TAM | Ui | In | M |
| 119 | <i>Daucus carota</i> L. | Apiaceae | H | A | CF | NAM | Fd | Cl | V |
| 120 | <i>Delonix regia</i> (Hook.) Raf. | Fabaceae | T | P | AR | TAF | Or | Cl | Or |
| 121 | <i>Dendrocalamus strictus</i> (Roxb.) Nees | Poaceae | S | P | F | AS | Ui | Nt | V |
| 122 | <i>Dentella repens</i> (L.) J.R.Forst. & G.Forst. | Rubiaceae | H | A | RB | AU | UI | Nt | Nk |
| 123 | <i>Dicoma tomentosa</i> Cass. | Asteraceae | H | A | W | TAM | Ui | In | M |
| 124 | <i>Digera muricata</i> (L.) Mart. | Amaranthaceae | H | A | CF | NAM | Ui | In | V |
| 125 | <i>Digitaria longiflora</i> (Retz.) Pers. | Poaceae | H | P | RB | TAF | Ui | Nt | Nk |
| 126 | <i>Dinebra retroflexa</i> (Vahl) Panz. | Poaceae | H | A | CF | TAM | Ui | Nt | NK |
| 127 | <i>Dioscorea bulbifera</i> L. | Dioscoreaceae | C | P | F | AS | M | Nt | V |
| 128 | <i>Duranta erecta</i> L. | Verbenaceae | S | P | CF | TAM | Or | Ca/Nt | Or |
| 129 | <i>Echinochloa colona</i> (L.) Link | Poaceae | H | A | W | EU | Fo | Nt | Fo |
| 130 | <i>Echinochloa crus-galli</i> (L.) P.Beauv. | Poaceae | H | A | CF | SAM | Fo | Nt | Fo |
| 131 | <i>Echinops echinatus</i> Roxb. | Asteraceae | H | A | W | TAF | Ui | Nt | M |
| 132 | <i>Eclipta prostrata</i> (L.) L. | Asteraceae | H | A | CF | TAM | Ui | In | M |
| 133 | <i>Eichhornia crassipes</i> (Mart.) Solms | Pontederiaceae | H | P | AQ | TAM | Or | In | Nk |
| 134 | <i>Eleocharis atropurpurea</i> (Retz.) J.Presl & C.Presl | Cyperaceae | H | A | AQ | SAM | Ui | Nt | Nk |

| | Accepted name of species | Family | Habit | Longevity | Habitat | Nativity | Purpose of introduction | Invasive status | Uses |
|-----|---|----------------|-------|-----------|---------|----------|-------------------------|-----------------|------|
| 135 | <i>Emilia sonchifolia</i> (L.) DC. ex DC. | Asteraceae | H | A | RB | TAM | Ui | In | M |
| 136 | <i>Eragrostis papposa</i> (Desf. ex Roem. & Schult.) Steud. | Poaceae | H | A | W | TAF | Ui | Nt | Nk |
| 137 | <i>Eryngium foetidum</i> L. | Apiaceae | H | A | W | TAM | UI | Nt/In | V |
| 138 | <i>Eucalyptus citriodora</i> Hk | Myrtaceae | T | P | W | AU | PI | Cl | Ol |
| 139 | <i>Euphorbia heterophylla</i> L. | Euphorbiaceae | H | A | CF | TAM | Ui | In | Or |
| 140 | <i>Euphorbia hirta</i> L. | Euphorbiaceae | H | A | CF | TAM | Ui | In | M |
| 141 | <i>Euphorbia pulcherrima</i> Willd. ex Klotzsch | Euphorbiaceae | S | P | W | MX | Or | Ca/Nt | M |
| 142 | <i>Euphorbia thymifolia</i> L | Euphorbiaceae | H | A | W | SAM | Ui | In | Nk |
| 143 | <i>Euphorbia tirucalli</i> L. | Euphorbiaceae | S | P | AR | TAM | Ui | Ca/Nt | M |
| 144 | <i>Euphorbia umbellata</i> (Pax) Bruyns. | Euphorbiaceae | S | P | W | TAM | Ui | In | M |
| 145 | <i>Ficus carica</i> L. | Moraceae | T | P | CF | EU | Fd | Nt | Ef |
| 146 | <i>Fimbristylis dichotoma</i> (L.) Vahl | Cyperaceae | H | A | RB | AS | Ui | Nt | Nk |
| 147 | <i>Foeniculum vulgare</i> Mill. | Apiaceae | H | A | CF | MR | Fd | Cl | Es |
| 148 | <i>Galinsoga parviflora</i> Cav. | Asteraceae | H | A | RB | TAM | Ui | In | M |
| 149 | <i>Glossocardia bosvallia</i> (L.f.) DC. | Asteraceae | H | A | F | WI | Ui | Nt | V |
| 150 | <i>Gnaphalium polycaulon</i> Pers. | Asteraceae | H | A | RB | TAM | Ui | In | Nk |
| 151 | <i>Gomphrena celosioides</i> Mart. | Amaranthaceae | H | A | W | TAM | Ui | Nt | Nk |
| 152 | <i>Gomphrena globosa</i> L. | Amaranthaceae | H | A | CF | TAM | Ui | In | Nk |
| 153 | <i>Grangea maderaspatana</i> (L.) Poir. | Asteraceae | H | A | RB | SAM | Ui | In | M |
| 154 | <i>Grevillea robusta</i> A.Cunn. ex R.Br. | Proteaceae | T | P | CF | AU | PI | Cl | W |
| 155 | <i>Guizotia abyssinica</i> (L.f.) Cass. | Asteraceae | H | A | CF | TAF | Ui | Cl | Ol |
| 156 | <i>Hamelia patens</i> Jacq | Rubiaceae | S | P | AR | BR | Or | Cl | Or |
| 157 | <i>Harrisia bonplandii</i> (Parm.) Britton & Rose | Cactaceae | S | P | W | SAM | Ui | Nt | Nk |
| 158 | <i>Helianthus annuus</i> L. | Asteraceae | H | A | CF | NAM | Ui | Cl | Ol |
| 159 | <i>Hibiscus cannabinus</i> L. | Malvaceae | S | P | CF | SAM | Ui | In | V |
| 160 | <i>Hibiscus rosa-sinensis</i> L. | Malvaceae | S | P | CF | AS | Or | Ca | Or |
| 161 | <i>Hibiscus sabdariffa</i> L. | Malvaceae | H | P | CF | SAM | Ui | Ca/Nt | V |
| 162 | <i>Hibiscus trionum</i> L. | Malvaceae | H | P | W | TAF | Ui | Nt | Nk |
| 163 | <i>Hyptis suaveolens</i> (L.) Poit. | Lamiaceae | H | P | AR | SAM | Ui | In | Nk |
| 164 | <i>Impatiens balsamina</i> L. | Balsaminaceae | H | A | RB | TAM | Or | Cl | Or |
| 165 | <i>Imperata cylindrica</i> (L.) Raeusch. | Poaceae | H | P | W | TAM | Ui | Nt | R |
| 166 | <i>Indigofera linifolia</i> (L.f.) Retz. | Fabaceae | H | A | RB | SAM | Ui | Nt | M |
| 167 | <i>Indigofera linnaei</i> Ali | Fabaceae | H | A | F | TAF | Ui | In | Nk |
| 168 | <i>Ipomoea alba</i> L. | Convolvulaceae | C | A | W | MX | Ui | Cl | Nk |
| 169 | <i>Ipomoea batatas</i> (L.) Lam. | Convolvulaceae | C | A | CF | BR | Fd | Cl | V |
| 170 | <i>Ipomoea cairica</i> (L.) Sweet | Convolvulaceae | C | A | W | TAF | Ui | Nt | Nk |
| 171 | <i>Ipomoea eriocarpa</i> R. Br. | Convolvulaceae | C | A | W | TAF | Or | Ca/Nt | Nk |
| 172 | <i>Ipomoea hederifolia</i> L. | Convolvulaceae | C | A | F | TAM | Ui | Nt | M |
| 173 | <i>Ipomoea nil</i> (L.) Roth | Convolvulaceae | C | A | W | NAM | Ui | In | Nk |
| 174 | <i>Ipomoea obscura</i> (L.) Ker Gawl. | Convolvulaceae | C | P | W | TAF | Ui | In | M |
| 175 | <i>Jatropha curcas</i> L. | Euphorbiaceae | S | P | AR | TAM | Or | Nt | Bf |
| 176 | <i>Jatropha gossypifolia</i> L. | Euphorbiaceae | S | P | W | TAM | Ui | Ca/Nt | Bf |
| 177 | <i>Kigelia pinnata</i> DC | Bignoniaceae | T | P | F | TAF | Or | Ca/Nt | Nk |
| 178 | <i>Lablab purpureus</i> (L.) Sweet | Fabaceae | H | A | CF | TAF | Fd | Cl | Ef |
| 179 | <i>Lagascea mollis</i> Cav. | Asteraceae | H | A | CF | TAM | Ui | In | Nk |
| 180 | <i>Lagenaria siceraria</i> (Molina) Standl. | Cucurbitaceae | C | A | AR | TAF | Ui | Nt | V |

| | Accepted name of species | Family | Habit | Longevity | Habitat | Nativity | Purpose of introduction | Invasive status | Uses |
|-----|--|-----------------|-------|-----------|---------|----------|-------------------------|-----------------|------|
| 181 | <i>Lantana camara</i> L. | Verbenaceae | S | P | F | TAM | Or | In | M |
| 182 | <i>Lawsonia inermis</i> L. | Lythraceae | S | P | W | TAF | Ui | Nt | M |
| 183 | <i>Leonotis nepetifolia</i> (L.) R.Br. | Lamiaceae | H | A | W | TAF | Ui | In | M |
| 184 | <i>Linum usitatissimum</i> L. | Linaceae | H | A | CF | EU | Fd | Cl | Es |
| 185 | <i>Ludwigia adscendens</i> (L.) H.Hara | Onagraceae | H | A | AQ | TAM | Ui | Nt | Nk |
| 186 | <i>Ludwigia octovalvis</i> (Jacq.) P.H.Raven | Onagraceae | H | A | RB | TAF | Ui | Nt | M |
| 187 | <i>Ludwigia perennis</i> L. | Onagraceae | H | A | RB | TAF | Ui | Nt | M |
| 188 | <i>Macrotyloma uniflorum</i> (Lam.) Verdc. | Fabaceae | H | A | CF | TAF | Fd | Cl | V |
| 189 | <i>Malvastrum coromandelianum</i> (L.) Garcke | Malvaceae | H | A | W | TAM | Ui | In | M |
| 190 | <i>Manihot esculenta</i> Crantz. | Euphorbiaceae | T | P | CF | SAM | Fd | Cl | V |
| 191 | <i>Manihot glaziovii</i> Muell. Arg. | Euphorbiaceae | T | P | CF | BR | Ui | Ca/Nt | Nk |
| 192 | <i>Manilkara zapota</i> (L.) P.Royen | Sapotaceae | S | P | CF | TAM | Ht | Cl | Ef |
| 193 | <i>Martynia annua</i> L. | Martyniaceae | H | P | W | NAM | Or | In | M |
| 194 | <i>Mecardonia procumbens</i> (Mill.) Small | Plantaginaceae | H | A | W | TAM | Ui | In | Nk |
| 195 | <i>Melia azedarach</i> L. | Meliaceae | T | P | AR | AS | M | Nt | W |
| 196 | <i>Melochia corchorifolia</i> L. | Sterculiaceae | H | P | F | TAM | Ui | In | V |
| 197 | <i>Mentha arvensis</i> L. | Lamiaceae | H | A | W | AS | Ui | Ca/Nt | M |
| 198 | <i>Merremia gangetica</i> Cufod. | Convolvulaceae | H | A | W | TAF | Ui | Nt | M |
| 199 | <i>Millingtonia hortensis</i> L. f. | Bignoniaceae | T | P | AR | AS | Ui | Ca/Nt | Or |
| 200 | <i>Mimosa pudica</i> L. | Mimosaceae | H | P | CF | BR | Ui | In | M |
| 201 | <i>Mirabilis jalapa</i> L. | Nyctaginaceae | H | A | W | SAM | Or | Nt | Or |
| 202 | <i>Monochoria vaginalis</i> (Burm.f.) C.Presl. | Pontederiaceae | H | P | RB | TAM | Ui | In | M |
| 203 | <i>Moringa oleifera</i> Lam. | Moringaceae | T | P | CF | NAM | Ht | Ca/Nt | V |
| 204 | <i>Morus alba</i> L. | Moraceae | S | P | CF | AS | Ht | Nt | Ef |
| 205 | <i>Muntingia calabura</i> L. | Muntingiaceae | T | P | AR | TAM | Or | Cl | Ef |
| 206 | <i>Mussaenda frondosa</i> L. | Rubiaceae | S | P | F | TAF | Ui | Cl | M |
| 207 | <i>Nerium oleander</i> L. | Apocynaceae | S | P | CF | EU | Or | Ca/Nt | Or |
| 208 | <i>Nicandra physalodes</i> (L.) Gaertn. | Solanaceae | H | A | W | SAM | Or | Ca | M |
| 209 | <i>Ocimum americanum</i> L. | Lamiaceae | H | A | W | TAM | Ui | In | M |
| 210 | <i>Opuntia ficus-indica</i> (L.) Mill. | Cactaceae | S | P | F | NAM | Ui | Nt/In | Ef |
| 211 | <i>Opuntia stricta</i> Haw. Var. <i>dillenii</i> (Ker Gawl.) | Cactaceae | S | P | F | TAM | Ui | Ca/Nt | Ef |
| 212 | <i>Oxalis corniculata</i> L. | Oxalidaceae | H | P | CF | EU | Ui | In | M |
| 213 | <i>Oxalis latifolia</i> Kunth | Oxalidaceae | H | A | W | BR | Ui | In | V |
| 214 | <i>Pandanus odorifer</i> (Forssk.) Kuntze | Pandanaceae | S | P | RB | SEA | Ui | Cl | M |
| 215 | <i>Parthenium hysterophorus</i> L. | Asteraceae | H | A | W | TAM | Ui | In | Nk |
| 216 | <i>Passiflora foetida</i> L. | Passifloraceae | C | P | W | SAM | Or | Cl | Or |
| 217 | <i>Passiflora subpeltata</i> Ortega | Passifloraceae | C | A | W | TAM | Ui | Nt | Nk |
| 218 | <i>Peltoperum pterocarpum</i> (DC.) Backer ex K. Heyne | Caesalpiniaceae | T | P | AR | AS | Ui | Ca/Nt | W |
| 219 | <i>Persicaria hydropiper</i> (L.) Delarbre | Polygonaceae | H | P | RB | EU | Fd | In | Nk |
| 220 | <i>Phaseolus vulgaris</i> L. | Fabaceae | H | A | CF | SAM | Fd | Cl | Es |
| 221 | <i>Phoenix sylvestris</i> (L.) Roxb. | Arecaceae | T | P | RB | TAM | Ui | Ca/Nt | Ef |
| 222 | <i>Phyla nodiflora</i> (L.) Greene | Verbenaceae | H | A | AQ | SAM | Ui | Ca/Nt | Nk |
| 223 | <i>Phyllanthus acidus</i> (L.) Skeels | Phyllanthaceae | T | P | AR | BR | Ui | Cl | Ef |
| 224 | <i>Phyllanthus amarus</i> Schumach. & Thonn. | Phyllanthaceae | H | A | W | TAM | Ui | Nt | M |
| 225 | <i>Physalis minima</i> L. | Solanaceae | H | A | W | NAM | M | In | Ef |
| 226 | <i>Pistia stratiotes</i> L. | Araceae | H | P | AQ | TAM | Ui | In | M |

| | Accepted name of species | Family | Habit | Longevity | Habitat | Nativity | Purpose of introduction | Invasive status | Uses |
|-----|--|------------------|-------|-----------|---------|----------|-------------------------|-----------------|------|
| 227 | <i>Pisum sativum</i> L. | Fabaceae | H | A | CF | TAM | Ht | Cl | Es |
| 228 | <i>Pithecellobium dulce</i> (Roxb.) Benth. | Mimosaceae | T | P | W | TAM | Ui | Nt | Ef |
| 229 | <i>Plumbago zeylanica</i> L. | Plumbaginaceae | S | P | W | TAF | Or | Cl | M |
| 230 | <i>Plumeria alba</i> L. | Apocynaceae | T | P | W | TAM | Or | Cl | Or |
| 231 | <i>Portulaca oleracea</i> L | Portulacaceae | H | A | W | SAM | Fd | In | V |
| 232 | <i>Portulaca pilosa</i> L. | Portulacaceae | H | A | W | SAM | Or | In | M |
| 233 | <i>Portulaca quadrifida</i> L. | Portulacaceae | H | A | W | TAM | Ui | In | M |
| 234 | <i>Potamogeton nodosus</i> Poir. | Potamogetonaceae | H | P | AQ | TAM | Ui | Nt | V |
| 235 | <i>Prosopis juliflora</i> (S.w.) DC | Mimosaceae | T | P | W | TAM | UI | Nt | M |
| 236 | <i>Psidium guava</i> L. | Myrtaceae | S | P | CF | SAM | Ht | Nt | Ef |
| 237 | <i>Punica granatum</i> L. | Lythraceae | T | P | CF | AS | Ht | Cl | Ef |
| 238 | <i>Pyrostegia venusta</i> (Ker Gawl.) Miers | Bignoniaceae | C | P | AR | BR | Or | Cl | Or |
| 239 | <i>Raphanus sativus</i> L. | Brassicaceae | H | A | CF | TAF | Fd | Cl | V |
| 240 | <i>Ricinus communis</i> L | Euphorbiaceae | S | A | W | TAF | Fd | In | Ol |
| 241 | <i>Rosa multiflora</i> Thunb. | Rosaceae | S | P | RB | AS | Or | Ca/Nt | M |
| 242 | <i>Rotala densiflora</i> (Roth) Koehne | Lythraceae | H | A | RB | AS | Ui | Nt | M |
| 243 | <i>Rubia cordifolia</i> L. | Rubiaceae | H | P | F | TAF | Ui | Nt | M |
| 244 | <i>Rubus ellipticus</i> Smith | Rosaceae | S | P | RB | TAM | Ui | Nt | Ef |
| 245 | <i>Ruellia prostrata</i> Poir. | Acanthaceae | H | A | W | TAF | Ui | In | M |
| 246 | <i>Ruta graveolens</i> L. | Rutaceae | H | A | W | MR | M | Cl | M |
| 247 | <i>Saccharum spontaneum</i> L. | Poaceae | S | P | RB | SEA | Ui | In | Fo |
| 248 | <i>Salvia coccinea</i> Buc'hoz ex EtL. | Lamiaceae | H | A | W | SAM | UI | Ca | Nk |
| 249 | <i>Scoparia dulcis</i> L. | Plantaginaceae | H | A | RB | TAM | M | In | Fo |
| 250 | <i>Sechium edule</i> (Jacq.) Sw. | Cucurbitaceae | C | A | CF | TAM | Ui | Cl | V |
| 251 | <i>Senna alata</i> (L.) Roxb. | Caesalpiniaceae | S | A | W | WI | Ui | In | M |
| 252 | <i>Senna occidentalis</i> (L.) Link | Caesalpiniaceae | S | P | W | SAM | Ui | In | M |
| 253 | <i>Senna sophra</i> (L.) Roxb. | Caesalpiniaceae | H | A | AR | WI | Ui | Nt/In | M |
| 254 | <i>Senna surattensis</i> (Burm.f.) H.S.Irwin & Barneby | Caesalpiniaceae | T | P | F | SEA | Ui | Cl | Nk |
| 255 | <i>Senna tora</i> (L.) Roxb. | Caesalpiniaceae | H | A | W | SAM | Ui | In | M |
| 256 | <i>Sesamum indicum</i> L. | Pedaliaceae | H | A | CF | TAF | Ui | Cl | Es |
| 257 | <i>Sesbania sesban</i> (L.) | Fabaceae | T | P | W | TAF | Ui | Nt | V |
| 258 | <i>Setaria italica</i> (L.) P.Beauv. | Poaceae | H | A | CF | TAF | Fo | In | Fo |
| 259 | <i>Sida acuta</i> Burm.f. | Malvaceae | H | A | W | TAM | Ui | Nt | M |
| 260 | <i>Sida cordata</i> (Burm. f.) Waalk. | Malvaceae | H | A | AR | SAM | Ui | Nt | M |
| 261 | <i>Siegesbeckia orientalis</i> L. | Asteraceae | H | A | AR | TAF | Ui | Ca/Nt | Nk |
| 262 | <i>Simarouba glauca</i> DC. | Simaroubaceae | T | P | AR | SAM | PI | Cl | M |
| 263 | <i>Solanum americanum</i> Mill. | Solanaceae | H | A | CF | TAM | Ui | In | V |
| 264 | <i>Solanum erianthum</i> D.Don | Solanaceae | H | P | F | TAM | Ui | In | M |
| 265 | <i>Solanum lycopersicum</i> L. | Solanaceae | H | P | CF | TAM | Ui | In | V |
| 266 | <i>Solanum melongena</i> L. | Solanaceae | H | A | CF | TAF | Fd | Cl | V |
| 267 | <i>Solanum pimpinellifolium</i> L. | Solanaceae | H | A | W | SAM | Ui | Cl | V |
| 268 | <i>Solanum seaforthianum</i> Andrews | Solanaceae | C | P | W | BR | Ui | In | Nk |
| 269 | <i>Solanum torvum</i> Sw. | Solanaceae | S | P | AR | WI | Ui | In | M |
| 270 | <i>Solanum tuberosum</i> L. | Solanaceae | H | P | CF | SAM | Fd | Cl | V |
| 271 | <i>Sonchus oleraceus</i> (L.) L. | Asteraceae | H | A | AR | MR | Ui | In | Nk |
| 272 | <i>Sonchus wightianus</i> DC. | Asteraceae | H | A | W | EU | Ui | Nt/In | M |

| | Accepted name of species | Family | Habit | Longevity | Habitat | Nativity | Purpose of introduction | Invasive status | Uses |
|-----|---|------------------|-------|-----------|---------|----------|-------------------------|-----------------|-------|
| 273 | <i>Spathodea campanulata</i> Beauv. | Bignoniaceae | T | P | AR | TAF | Or | Ca/Nt | Or |
| 274 | <i>Spermacoce hispida</i> L. | Rubiaceae | H | A | W | TAM | Ui | In | M |
| 275 | <i>Sphagneticola calendulacea</i> (L.) Pruski | Asteraceae | H | A | W | AU | Ui | Nt | Or |
| 276 | <i>Sporobolus diander</i> (Retz.) P. Beauv. | Poaceae | H | A | W | AS | Ui | In | Fo |
| 277 | <i>Stachytarpheta jamaicensis</i> (L.) Vahl | Verbenaceae | S | P | F | TAM | Ui | In | M |
| 278 | <i>Stachytarpheta mutabilis</i> (Jacq.) Vahl. | Verbenaceae | S | P | W | SAM | Ui | Ca/Nt | Nk |
| 279 | <i>Stylosanthes fruticosa</i> (Retz.) Alston | Fabaceae | H | P | W | TAM | Ui | In | Fo |
| 280 | <i>Swietenia mahagoni</i> (L.) Jack. | Meliaceae | T | P | AR | WI | Ui | Nt | M |
| 281 | <i>Synadenium grantii</i> Hook. f. | Euphorbiaceae | S | P | W | TAM | Or | In | M |
| 282 | <i>Synedrella nodiflora</i> (L.) Gaertn. | Asteraceae | H | A | W | WI | Ui | In | Nk |
| 283 | <i>Tabebuia aurea</i> (Silva Manso) Benth. & Hook.f. ex S.Moore | Bignoniaceae | T | P | AR | TAM | Or | Ca/Nt | Or |
| 284 | <i>Tabebuia rosea</i> (Bertol.) Bertero ex A.DC. | Bignoniaceae | T | P | AR | TAM | Or | Cl | Or |
| 285 | <i>Tagetes erecta</i> L. | Asteraceae | H | P | CF | TAM | Or | Cl | Or |
| 286 | <i>Tagetes patula</i> L. | Asteraceae | H | A | W | MX | Or | Ca | Or |
| 287 | <i>Talinum portulacifolium</i> (Forssk.) Asch. ex Schweinf. | Portulacaceae | H | A | W | TAM | Ui | Nt/In | V |
| 288 | <i>Tamarindus indica</i> L. | Fabaceae | T | P | AR | TAF | Ht | Ca/Nt | Ef |
| 289 | <i>Tecoma capensis</i> (Thunb.) Lindl. | Bignoniaceae | S | P | CF | EU | Or | Cl | Or |
| 290 | <i>Tecoma gaudichandi</i> DC. | Bignoniaceae | S | P | AR | SAM | Or | Cl | Or |
| 291 | <i>Tecoma stans</i> (L.) Juss. ex Kunth | Bignoniaceae | T | P | AR | TAM | Or | Cl | Or |
| 292 | <i>Thunbergia alata</i> Bojer ex Sims | Acanthaceae | C | P | AR | TAF | Or | In | Or |
| 293 | <i>Tithonia diversifolia</i> (Hemsl.) A.Gray | Asteraceae | S | A | W | MX | Or | In | Or |
| 294 | <i>Torenia fournieri</i> Linden ex E. Fourn. | Linderniaceae | H | P | W | AU | Ui | In | Or |
| 295 | <i>Tradescantia spathacea</i> Sw. | Commelinaceae | H | A | W | TAM | Ui | Cl | Or |
| 296 | <i>Trapa natans</i> L. | Lythraceae | H | P | AQ | EU | Fd | In | Ef |
| 297 | <i>Tribulus terrestris</i> L. | Zygophyllaceae | H | P | W | TAM | Ui | In | M |
| 298 | <i>Tridax procumbens</i> (L.) L. | Asteraceae | H | P | W | TAM | Ui | In | M |
| 299 | <i>Trigonella foenum-graecum</i> L. | Fabaceae | H | A | CF | MR | Fd | Cl | Es |
| 300 | <i>Triumfetta rhomboidea</i> Jacq. | Malvaceae | H | A | W | TAM | Ui | In | M |
| 301 | <i>Typha angustifolia</i> L. | Typhaceae | H | P | RB | EU | Ui | In | Fo |
| 302 | <i>Typha domingensis</i> Pers | Typhaceae | H | P | AQ | SAM | Ui | In | Nk |
| 303 | <i>Urena lobata</i> L. | Malvaceae | S | P | AR | TAF | Ui | Ca/Nt | M |
| 304 | <i>Urochloa panicoides</i> P. Beauv. | Poaceae | H | A | W | TAF | Ui | In | Fo |
| 305 | <i>Vallisneria spiralis</i> L. | Hydrocharitaceae | H | A | AQ | MR | Ui | Nt/In | Nk |
| 306 | <i>Vigna trilobata</i> (L.) Verdc. | Fabaceae | C | A | W | TAF | UI | Nt | M |
| 307 | <i>Vigna umbellata</i> (Thunb.) Ohwi & H.Ohashi | Fabaceae | C | A | CF | SEA | Ui | Nt/In | Es |
| 308 | <i>Vigna unguiculata</i> (L.) Walp. | Fabaceae | H | A | CF | TAM | Fd | Cl | V |
| 309 | <i>Vitex negundo</i> L. | Verbenaceae | S | P | W | AS | UI | Ca/Nt | M |
| 310 | <i>Waltheria indica</i> L. | Sterculiaceae | H | P | F | TAM | Ui | In | M |
| 311 | <i>Xanthium strumarium</i> L. | Asteraceae | H | A | AR | TAM | Ui | In | M |
| 312 | <i>Zea mays</i> L. | Poaceae | H | A | CF | SAM | Fd | Cl | Fo, V |

Habit: H—Herb | S—Shrub | C—Climber | T—Tree | **Longevity:** A—Annual | P—Perennial | **Habitat:** W—Wasteland | CF—Cultivated fields | RB—River or pond banks | F—Forests | AR—Roadsides | AQ—Aquatic | P—Parasite | **Nativity:** AS—Tropical Asia | AU—Australia | BR—Brazil | EU—Europe | MG—Madagascar; MR—Mediterranean region | MX—Mexico | NAM—North America | SAM—South America | SEA—South East Asia (Including Malaysia, Philippines & Indonesia); TAF—Tropical Africa | TAM—Tropical America | WI—West Indies | **Mode of introduction to India:** Fd—Food | Fo—Fodder | M—Medicine | O—Ornamental | PI—Plantation | Ht—Horticultural; Ui—Unintentional | **Status:** Cl—Cultivated | Ca—Casual | Nt—Naturalized | In—Invasive | Ca/Nt—Casual or Naturalized | Nt/In—Naturalized or Invasive | **Uses:** Bf—Biofuel | Br—Beverages | Ef—Edible fruit | Es—Edible seed | Fo—Fodder | M—Medicinal | Nk—Not known | Ol—Oil | Or—Ornamental; R—Rope making | V—Vegetable | W—Wood.

cylindrica, *Opuntia stricta*, *Clidemia hirta*, *Lantana camara*, *Chromolaena odorata*, and *Rubus ellipticus*. Pysek et al. (2017) have identified 11 alien plant species that occur on one-third or more of the globe in terms of the number of regions where they are naturalized, and on at least 35% of the Earth's land surface. Of these, eight plant species are widely distributed in Hassan district: *Bidens pilosa*, *Chenopodium album*, *Datura stramonium*, *Echinochloa crus-galli*, *Oxalis corniculata*, *Portulaca oleracea*, *Ricinus communis*, and *Sonchus oleraceus*. The impact of these species on indigenous flora and invading ecosystems, however, has yet to be studied. The distribution of alien plant species was most abundant in wastelands (34%), followed by cultivated fields (30%), roadsides (14%), river or pond banks (9%), forests (8%), and aquatic systems (4%) (Figure 3). This pattern could be caused by the relative degree of disturbance in various environments, as well as other abiotic and biotic factors. Disturbance alters the physical environment creating open regions and disturbed environmental factors, such as, elevated soil nitrate and increased light and temperature changes, boost seed germination for many species, including exotics. This could allow alien species to establish themselves in ecosystems. Several researchers have discovered that the species composition after disturbance is reasonably predicted based on the seed bank before disturbance. As a result, sampling the pre-disturbance seed bank can provide insight into whether exotics will become abundant at a site in the event of a predicted disturbance (D'antonio & Meyerson 2002).

We categorized the origin of the reported invasive species into 12 regions, of which tropical America was found to be the origin for 36% (113 species), followed by tropical Africa 15% (48 species), South America 13% (41 species), and tropical Asia (28 species). The other regions, contribute 2–5 % each to the overall alien flora (Figure 4). The possible explanation for the maximum proportion of species from tropical America can be the higher propagule pressure from different countries, such as Brazil and Mexico, to India via historical trade routes through the human agency of European colonisers and traders, and more or less matching of similar tropical climate (Khuroo et al. 2012). Considering that 36% of species originate in the Americas, the findings of this study are comparable to those reported for China, where 58% of species originate in the Americas (Wu et al. 2010). However, compared to the current study, the percentage of American species in the alien flora of Europe is lower at 34.8% (Lambdon et al. 2008). Because tropical climates have a higher impact on India and China

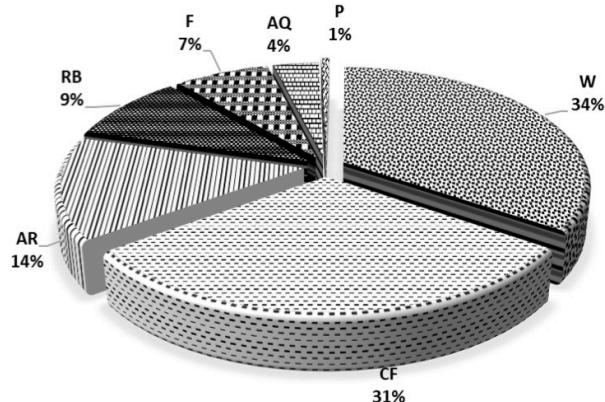


Figure 3. Habitat wise distribution of alien species in Hassan district.
W—Wasteland | CF—Cultivated fields | RB—River or pond banks | F—Forests | AR—Roadsides | AQ—Aquatic | P—Parasite.

Table 2. Relative contribution of alien species in the top 10 alien species rich families in Hassan district of Karnataka.

| | Family | Alien species | Total species in Hassan district | Alien plants (%) |
|----|----------------|---------------|----------------------------------|------------------|
| 1 | Amaranthaceae | 17 | 20 | 85 |
| 2 | Solanaceae | 15 | 18 | 83.3 |
| 3 | Asteraceae | 36 | 76 | 47.3 |
| 4 | Convolvulaceae | 9 | 19 | 47.3 |
| 5 | Malvaceae | 12 | 27 | 44.4 |
| 6 | Caesalpinaeae | 8 | 23 | 34.7 |
| 7 | Fabaceae | 21 | 97 | 33.3 |
| 8 | Euphorbiaceae | 14 | 51 | 27.4 |
| 9 | Apocynaceae | 8 | 24 | 25 |
| 10 | Poaceae | 16 | 140 | 11.4 |

than on Europe, this distinct pattern can be explained.

Some alien plant species, 36% of those listed for Hassan district, are used for medicinal purposes, followed by vegetables (16%), ornamentals (13%), edible fruits (8%), fodder (4%), timber (2%), and biofuel (1%). A large number of alien plant species benefit Indian agriculture, forestry, and pharmaceutical industries, as well as the Indian medical system (Ayurveda) (Shiddamallaya et al. 2010). Ornamental plants are an important component of the urban environment, as well as a substantial source of invasive species as a result of escapes from private or public gardens (Pyek & Chytr 2014; Pergl et al. 2016). Many taxa first escape and spread in spatially constrained areas around gardens, before spreading and colonising more distant vegetation. The combined impacts of local popularity of a specific taxon, regardless of invasion status, adequate natural & cultural conditions, abundant

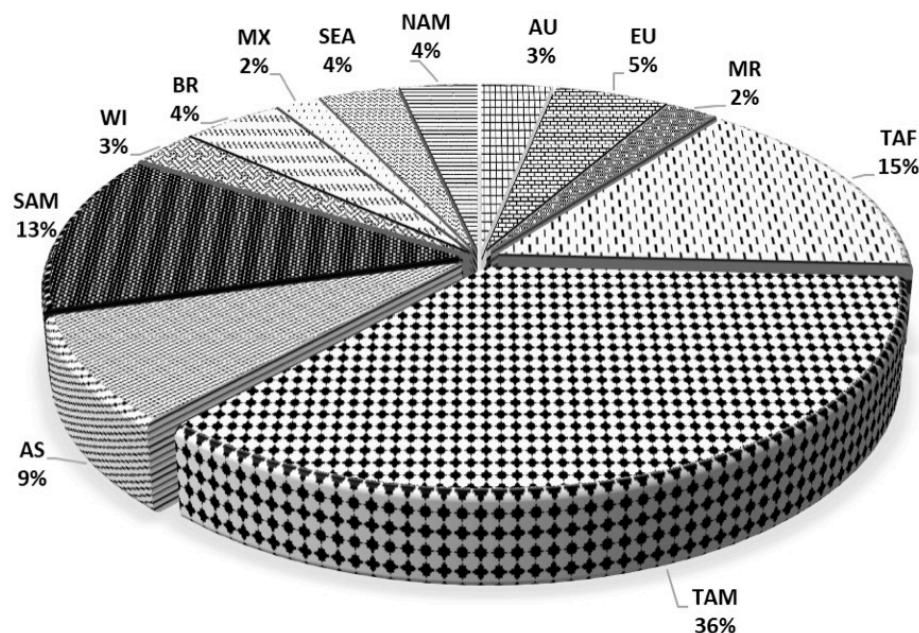


Figure 4. The Nativity of alien species of Hassan District.

SAM—South America | SEA—South East Asia | BR—Brazil | TAM—Tropical America | EU—Europe | TAF—Tropical Africa | AS—Tropical Asia | AU—Australia | WI—West Indies | MR—Mediterranean region | MX—Mexico | NAM—North America.

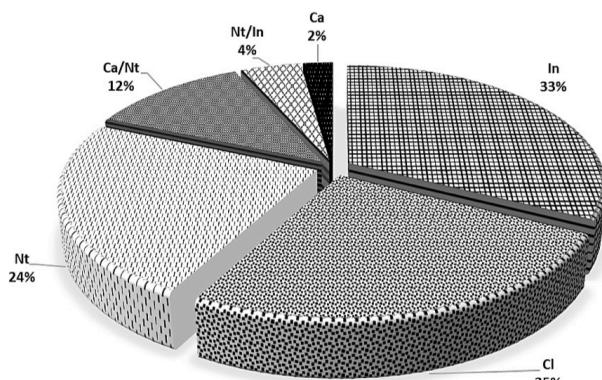


Figure 5. Invasive status of the alien species of Hassan district.
CI—Cultivated | Ca—Casual | Nt—Naturalized | In—Invasive | Ca/Nt—Casual or Naturalized | Nt/In—Naturalized or Invasive.

propagation in cultivation, and easy semi-spontaneous establishment in gardens may result in naturalisation foci (Petrik et al. 2019). Although the majority of alien species grown as garden ornamentals can only survive when planted under careful management, a significant proportion of them manage to escape and establish themselves outside of human control (Pergl et al. 2016). Pyek et al. (2012) found that 56% of the taxa in the Czech Republic's alien flora were recruited from escaping ornamental plants. Similarly, in the Karnataka district of Gadag, roughly 15% of alien species are employed as ornamentals (Kambhar & Kotresha 2011).

We found that 122 species were intentionally introduced, while the rest are unintentional introductions. The majority of species were introduced for ornamental purposes (47%), followed by food (30%), horticulture (10%), medicinal (9%), fodder (4%), and plantation (2%). The invasion status categorization of Hassan is represented in Figure 5.

Hassan district is the reservoir of rich flora, and is a significant segment of the global biodiversity hotspot of Western Ghats. Approximately 1,700 vascular plant species found in Hassan district accounts for 75% of the plant species of Karnataka state and 10% of India, which indicates the richness of biological diversity (Saldhana & Nicolson 1978). However, almost 18.4% of Hassan district flora comprises of alien species, which is higher than the 8% of alien plants in Western Ghats region of Karnataka and 6.5% alien species of Karnataka state (Ganeshaiah et al. 2002; Rao 2012). The majority of the alien species belong to the family Asteraceae, and it also contributed most of the exotic weed species in India (Singh et al. 2010; Kambhar & Kotresha 2011; Khuroo et al. 2012).

In concordance with the alien floras of Europe (Lambdon et al. 2008) and China (Wu et al. 2010), Asteraceae is the most species-rich family in the alien flora of India. At the global level, Pysek (1998) found these families to be having the majority of alien species. Studies on agricultural weeds concluded that, numerically, most

weeds come from the families Asteraceae (Heywood 1989). Notably, Asteraceae is amongst the largest family in terms of species richness (Rao 1994). Hence, the possibility of contributing more to alien species is also higher (Mack & Erneberg 2002). The introduction of alien plants for ornamental purpose is common across the globe and especially species belongs to the genera *Amaranthus*, *Cascabela*, *Euphorbia*, *Ipomoea*, and *Solanum* are some of the commonly preferred ornamental alien species reported in India (Khuroo et al. 2012). *Alternanthera philoxeroides* powerful aquatic pest has been found in the lakes, ponds, puddles and waterways was considered a highly invasive and spread throughout the country (Maheshwari 1965). *Lantana camara*, *Chromolaena odorata*, and *Hyptis suaveolens* were the most concerning alien invasive plant species in terms of rapid growth, higher density, and frequency in forest areas.

Within the forest, these species were so gregarious in their growth and most ecologically destructive invaders in the Western Ghats region (Muniappan & Viraktamath 1993). The escape of these species into nature, on the other hand, may have serious consequences. *Chromolaena odorata* is an invasive transformer species in the Old World (Richardson et al. 2000), owing to its lack of natural enemies. It prefers areas of natural or human-induced disturbance, but it can even infiltrate untouched terrain. Subsistence and commercial agriculture, including crops and plantations, grazing pastures, and silviculture, are all affected by *Chromolaena odorata*. Awanyo et al. (2011) mentioned that the highly invasive *Chromolaena odorata* grows aggressively and suppresses other vegetation by easily forming a thick cover in a very short time. In another study, the high allelopathic properties of this weed support its gaining dominance in vegetation and in replacing other aggressive invaders such as *Lantana camara* and *Imperata cylindrica* in Asia and Africa (Mandal & Joshi 2014). The most common species of invasives in cultivated areas were *Celosia argentea* and *Argemone mexicana*, which were so aggressive and opportunistic in invasion that they could even penetrate flourishing crops if regular weeding was neglected. *Ageratum conyzoides*, *Cassia tora*, *Emilia sonchifolia*, *Oxalis corniculata*, *Scoparia dulci*, *Sonchus oleraceus*, and *Tridax procumbens* are some of the other invasive plant species commonly found in cultivated fields that require constant weeding in practices and act as vectors for transmitting pathogens. Parasitic dodders (*Cuscuta* spp.) are becoming a severe concern in south Indian agroecosystems, and are increasingly being detected on

a wide range of plants across the country.

Ipomoea carnea, *Pistia stratiotes*, and *Eichhornia crassipes* have become a nuisance in aquatic ecosystems. They cause hindrance and block drainage and reduces the aesthetic value of open water bodies (Kambhar & Kotresha 2011). The invasion of *Eichhornia crassipes* into freshwater systems poses a threat to many human uses. Boating access, navigability, and recreation, as well as pipe systems for agriculture, industry, and municipal water supply, are the most direct impacts. Fish catchability and access to fishing grounds are also impacted. Furthermore, *Eichhornia crassipes* evapotranspiration rates can be higher than open-water evaporation rates. This can be a major issue in water-scarce places and small bodies of water. If it causes changes in fish community composition or modifies the catchability of fished species, it can have a significant impact on fishery (Villamagna & Murphy 2010).

Aside from the negative effects on native flora and the economy, certain alien species were useful to locals. Leafy vegetables included *Portulaca oleracea*, *Chenopodium album*, *Alternanthera sessilis*, *Amaranthus spinosus*, *Digera muricata*, and *Solanum americanum*. In its invaded area in India, *Prosopis juliflora* grows in forests, wastelands, and at the edges of crop fields, forming pure stands. Farmers retain trees in their fields because their crops grow better under them than in open fields, but they also provide fuel, fodder, charcoal, and lumber (Kaur et al. 2012). Invasive alien plant inventories are one of the most important components for assessing biodiversity and threats to endangered species, as well as providing source data for developing relevant indicators (Pyek et al. 2012; van Kleunen et al. 2015; Latombe et al. 2017). Identifying invasive alien plant species that pose prospective or future threats while they are still in the early stages of invasion is a serious prediction challenge (Lambdon et al. 2008). The findings of this study will raise awareness of invasive alien plants, and the release of this list will encourage more data collection so that the effects of these species can be minimized.

CONCLUSION

The present paper provides information on the status of alien plant species in Hassan district. It is revealed that over 18% of Hassan district flora comprises of alien species, which is higher than the 8% for the Western Ghats region of Karnataka and 6.5% of alien species in Karnataka state. A majority of the species are of South



Image 1. A—*Acanthospermum hispidum* DC | B—*Ageratum houstonianum* Mill. | C—*Alternanthera paronychioides* A.St.-Hil. | D—*Alternanthera philoxeroides* (Mart.) Griseb. | E—*Alternanthera pungens* Kunth | F—*Alternanthera sessilis* (L.) R.Br. ex DC. | G—*Amaranthus spinosus* L. | H—*Amaranthus viridis* L | I—*Argemone mexicana* L. | J—*Asclepias curassavica* L. | K—*Balanites aegyptiaca* (L.) Delile. | L—*Basella alba* L. © G M Prashanth Kumar.



Image 2. A—*Bixa orellana* L. | B—*Blainvillea acmella* (L.) Philipson | C—*Cardiospermum halicacabum* L. | D—*Cascabela thevetia* (L.) Lippold. | E—*elosia argentea* L. | F—*Chromolaena odorata* (L.) RM.King& H.Rob. | G—*Cleome viscosa* L. | H—*Clitoria ternatea* L. | I—*Corchorus aestuans* L. | J—*Cuscuta reflexa* Roxb. | K—*Datura metel* L. | L—*Dicoma tomentosa* Cass. © G M Prashanth Kumar.

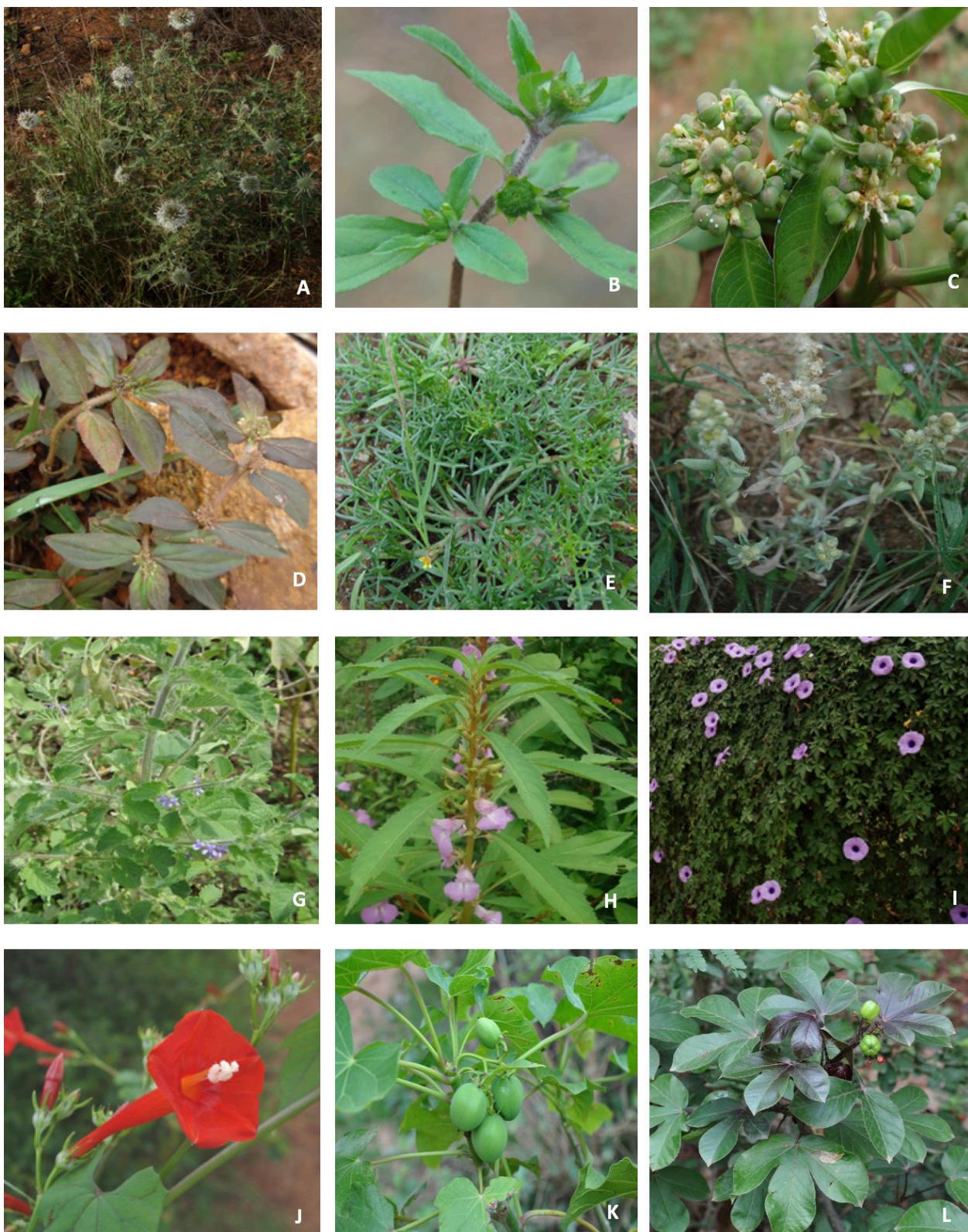


Image 3. A—*Echinops echinatus* Roxb. | B—*Eclipta prostrata* (L.) L. | C—*Euphorbia heterophylla* L. | D—*Euphorbia hirta* L. | E—*Glossocardia bosvallia* (L.f.) DC | F—*Gnaphalium polycaulon* Pers. | G—*Hyptis suaveolens* (L.) Poit. | H—*Impatiens balsamina* L. | I—*Ipomoea cairica* (L.) Sweet | J—*Ipomoea hederifolia* L. | K—*Jatropha curcas* L. | L—*Jatropha gossypifolia* L. © G M Prashanth Kumar.



Image 4. A—*Lantana camara* L. | B—*Malvastrum coromandelianum* (L.) Garcke | C—*Martynia annua* L. | D—*Oxalis corniculata* L. | E—*Pandanus odorifer* (Forssk.) Kuntze | F—*Passiflora foetida* L. | G—*Phyllanthus amarus* Schumach. & Thonn. | H—*Physalis minima* L. | I—*Pistia stratiotes* L. | J—*Pithecellobium dulce* (Roxb.) Benth. | K—*Portulaca oleracea* L. | L—*Prosopis juliflora* (S.w.) DC. © G M Prashanth Kumar.



Image 5. A—*Scoparia dulcis* L. | B—*Senna occidentalis* (L.) Link | C—*Senna tora* (L.) Roxb. | D—*Solanum americanum* Mill. | E—*Solanum seaforthianum* Andrews | F—*Solanum torvum* Sw. | G—*Sonchus oleraceus* (L.) L. | H—*Stachytarpheta jamaicensis* (L.) Vahl | I—*Stylosanthes fruticosa* (Retz.) Alston. | J—*Thunbergia alata* Bojer ex | K—*Tribulus terrestris* L. | L—*Typha angustifolia* L. © G M Prashanth Kumar.

American origin and have been introduced for ornamental purposes. Our study indicated that the extent and present share of alien species and their naturalization cannot be considered safe for native and endemic flora. This is especially true of Hassan district, which is part of the Western Ghats 'hotspot' belt and is globally designated for priority of conservational activities. As most forests of the Western Ghats are already badly affected by the invasion of alien plant species, the need for effective control must be emphasized. This compiled work will fill a significant information gap regarding alien species, and will aid in the development of informed monitoring and management strategies, always preserving site biodiversity and peoples' cultural diversity in mind, rather than simply the scale of bio-invasion.

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